

## Established Insecticide Target Site Mutations (ver 2.4)

Development of insecticide resistance can result from various mechanisms: increased ability for insecticide detoxification, decreased penetration/transport, or modification of the insecticide's target site. Selection pressure may lead to resistance from a single mechanism or combination of resistance mechanisms (ie. target-based and metabolic). Mutation of an amino acid residue (or multiple residues) at the target site can dramatically reduce sensitivity to an entire class of insecticides. This table provides a comprehensive list of established target site mutations associated with published cases of insecticide resistance.

IRAC MoA Group	Target Site	Affected Organisms	Mutation	Subunit	Mutation Common Name	Field Relevance	Literature References
1A	Acetylcholinesterase (Carbamates)	<i>Aphis gossypii</i>	S431F A302S, S431F				<a href="#">Andrews et al. (2004) Insect Mol Biol. 13:555</a> <a href="#">Toda et al. (2004) Insect Mol Biol. 13:549</a> <a href="#">Khajehali et al. (2010) Pest Manag Sci. 66:220</a> <a href="#">Carvalho et al. (2012) Pest Biochem Physiol. 104:143</a> <a href="#">Ilías et al. (2004) Insect Biochem Mol Biol. 48:17</a> <a href="#">Wu et al. (2015) Insect Biochem Mol Biol. 65:75</a> <a href="#">Badolo et al. (2015) Malar J. 14:477</a> <a href="#">Feng et al. (2015) Malar J. 14:470</a>
		<i>Tetranychus urticae</i>	A201S, T280A, F331C/Y/W, G328A F331Y			Yes	
		<i>Apolysmus lucorum</i>	A201S, T280A, F331C/Y/W, G328A A216S				
		<i>Anopheles gambiae</i>	G119S G119S				
1B	Acetylcholinesterase (Organophosphates)	<i>Aphis gossypii</i>	S431F, A302S				<a href="#">Andrews et al. (2004) Insect Mol Biol. 13:555</a> <a href="#">Toda et al. (2004) Insect Mol Biol. 13:549</a> <a href="#">Kakani et al. (2008) Insect Biochem Mol Biol. 38:781</a> <a href="#">Khajehali et al. (2010) Pest Manag Sci. 66:220</a> <a href="#">Carvalho et al. (2012) Pest Biochem Physiol. 104:143</a> <a href="#">Ilías et al. (2004) Insect Biochem Mol Biol. 48:17</a> <a href="#">Wu et al. (2015) Insect Biochem Mol Biol. 65:75</a>
		<i>Bactrocera oleae</i>	Δ3Q			Yes	
		<i>Tetranychus urticae</i>	A201S, T280A, F331C/Y/W, G328A F331Y				
		<i>Apolysmus lucorum</i>	A201S, T280A, F331C/Y/W, G328A A216S				
2	GABA-gated chloride channel	<i>Nilaparvata lugens</i>	R300Q			Yes	<a href="#">Zhang et al. (2016) Scientific Reports. doi:10.1038/srep32335</a>
		<i>Bemisia tabaci</i>					<a href="#">Anthony et al. (1995) Pest Biochem Physiol. 51:220</a>
		<i>Sogatella furcifera</i>					<a href="#">Nakao et al. (2010) Pest Biochem Phys. 97:262</a>
		<i>Laodelphax striatellus</i>	A302S/N	α	rdl	Yes	<a href="#">Nakao et al. (2011) J Econ Entomol. 104:646</a> <a href="#">Wang et al. (2016) J Econ Entomol. 109:334</a> <a href="#">French-Constant et al. (1993) Nature. 363:44</a> <a href="#">Le Goff et al. (2005) J Neurochem. 92:1295</a>
		<i>Plutella xylostella</i>				No	
		<i>Drosophila melanogaster</i>	A302S				
3	Sodium channel	<i>Drosophila simulans</i>	A302S (A301G), T350M				
		<i>Helicoverpa zea</i>	V421A/G, L1029H	IS6		Yes	<a href="#">Hopkins &amp; Pietrantonio. (2010) Insect Biochem Mol Biol. 40:385</a>
		<i>Cimex lectularius</i>	V419L, L925I	IS6		Yes	<a href="#">Yoon et al. (2008) J Med Entomol. 45:1092</a>
		<i>Heliothis virescens</i>	V421M	IS6		Yes	<a href="#">Park et al. (1997) Biochem Biophys Res Comm. 239:688</a>
		<i>Plutella xylostella</i>	M918I	IIS4-S6		Yes	<a href="#">Sonoda et al. (2012) Pest Biochem Physiol. 102:142</a>
		<i>Myzus persicae</i>	M918L, L932F	IIS4-S5		Yes	<a href="#">Fontaine et al. (2011) Pest Manag Sci. 67:881</a>
		<i>Thrips tabaci</i>	M918L	IIS4-S5		Yes	<a href="#">Wu et al. (2014) Pest Manag Sci. 70:977</a>
		<i>Musca domestica</i>	M918T, L1014F	IIS4-S6		No	<a href="#">Williamson et al. (1996) Molec Gen Genet. 252:51</a>
		<i>Tetranychus evansi</i>	M918T	IIS4-S5		Yes	<a href="#">Nyoni et al. (2011) Pest Manag Sci. 67:891</a>
		<i>Myzus persicae</i>	M918T	IIS4-S5		Yes	<a href="#">Eleftherianos et al. (2008) Bull Entomol Res. 98:183</a>
		<i>Thrips tabaci</i>	M918T, T929I, L1014F	IIS4-S6	super-kdr	Yes	<a href="#">Toda &amp; Morishita. (2009) J Econ Entomol. 102:2296</a>
		<i>Tuta absoluta</i>	M918T, T929I, L1014F	IIS4-S6	super-kdr	Yes	<a href="#">Haddi et al. (2012) Insect Biochem Mol Biol. 42:506</a>
		<i>Bemisia tabaci</i>	M918V, L925I	IIS4-S5		Yes	<a href="#">Morin et al. (2002) Insect Biochem Mol Biol. 32:1781</a>
		<i>Bemisia tabaci</i>	L925I, T929V	IIS5		Yes	<a href="#">Rodilakis et al. (2006) Pest Biochem Phys. 85:161</a>
		<i>Rhipicephalus microplus</i>	C190A	IIS5		Yes	<a href="#">Moman et al. (2009) Int J Parasit. 39:775</a>
		<i>Varroa destructor</i>	L925V	IIS		Yes	<a href="#">Gonzalez-Cabrer et al. (2013) PLoS One. 8:e62941</a>
		<i>Plutella xylostella</i>	T929V, L1014F	IIS4-S6		Yes	<a href="#">Sonoda et al. (2008) Insect Biochem Mol Biol. 38:883</a>
		<i>Ctenocephalides felis</i>	T929V, L1014F	IIS4-S6		Yes	<a href="#">Bass et al. (2014) Insect Biochem Mol Biol. 34:1305</a>
		<i>Pediculosis captis</i>	T929I, L932F	IIS4-S5		Yes	<a href="#">Lee et al. (2000) Pest Biochem Phys. 66:130</a>
		<i>Aedes aegypti</i>	I1011M, V1016G	IIS6		Yes	<a href="#">Saavedra-Rodrigues et al. (2007) Insect Mol Biol. 16:785</a>
		<i>Anopheles sinensis</i>	L1014F	IIS6	kdr	Yes	<a href="#">Kim et al. (2007) Pest Biochem Physiol. 87:54</a>
		<i>Plutella xylostella</i>	L1014F	IIS6	kdr	Yes	<a href="#">Schuler et al. (1998) Pest Biochem Phys. 59:169</a>
		<i>Blattella germanica</i>	L1014F	IIS6	kdr	No	<a href="#">Dong. (1997) Insect Biochem Mol Biol. 27:93</a>
		<i>Myzus persicae</i>	L1014F	IIS6	kdr	Yes	<a href="#">Martinez-Torres et al. (1999) Insect Mol Biol. 8:339</a>
		<i>Culex pipiens</i>	L1014F	IIS6	kdr	No	<a href="#">Martinez-Torres et al. (1999) Pest Sci. 55:1012</a>
		<i>Tecia solenivora</i>	L1014F	IIS6	kdr	Yes	<a href="#">Bacca et al. (2016) Pest Manag Sci. xx:xx</a>
		<i>Anopheles gambiae</i>	L1014F	IIS6	kdr	Yes	<a href="#">Hanson et al. (2000) Insect Mol Biol. 9:491</a>
		<i>Anopheles sacharovi</i>	L1014F/S	IIS6	kdr	Yes	<a href="#">Ulevap et al. (2002) J Med Entomol. 39:870</a>
		<i>Anopheles stephensi</i>	L1014F	IIS6	kdr	Yes	<a href="#">Enayati et al. (2003) Med Vet Entomol. 17:138</a>

		<i>Anopheles gambiae</i> <i>Anopheles arabiensis</i>	L1014F/S	IIS6	kdr	Yes	<a href="#">Verhaeghen et al. (2006) Malaria J. 5:16</a> <a href="#">Elang et al. (2006) Am J Trop Med Hyg. 74:795</a> <a href="#">Kanumaratne et al. (2007) Pest Biochem Phys. 88:108</a> <a href="#">Marshall et al. (2012) J Pest Sci. 37:169</a> <a href="#">Nauen et al. (2012) Pest Biochem Phys. 103:173</a> <a href="#">Foster et al. (2013) Pest Manag Sci. 70:1248</a> <a href="#">Zimmer et al. (2014) Pest Biochem Physiol. 108:1</a> <a href="#">Rinkevich et al. (2006) Insect Mol Biol. 15:157</a> <a href="#">Tan et al. (2012) J Med Entomol. 49:1012</a> <a href="#">Endensby et al. (2011) Bull Entomol Res. 101:393</a> <a href="#">Ilis et al. (2004) Insect Biochem Mol Biol. 48:17</a> <a href="#">Harris et al. (2010) Am J Trop Med Hyg. 83:277</a> <a href="#">Pasav et al. (2008) Med Vet Entomol. 22:82</a> <a href="#">He et al. (1999) Biochem Biophys Res Comm. 261:558</a> <a href="#">Ding et al. (2015) Pest Manag Sci. 71:266</a> <a href="#">Head et al. (1998) Insect Mol Biol. 7:191</a>
		<i>Anopheles subpictus</i>	L1014F	IIS6	kdr	Yes	
		<i>Aphis gossypii</i>	L1014F	IIS6	kdr	Yes	
		<i>Meligethes aeneus</i>	L1014F	IIS6	kdr	Yes	
		<i>Sitobion avenae</i>	L1014F	IIS6	kdr	Yes	
		<i>Psylliodes chrysoccephala</i>	L1014F	IIS6	kdr	Yes	
		<i>Musca domestica</i>	L1014F/H	IIS6	kdr	No	
		<i>Anopheles sinensis</i>	L1014F/S/W	IIS6	kdr	Yes	
		<i>Plutella xylostella</i>	F1020S	IIS6		Yes	
		<i>Tetranychus urticae</i>	A1215D, F1538I	IIS6-IIIS6		Yes	
		<i>Aedes aegypti</i>	F1534C	IIIS6		Yes	
		<i>Sarcophaga scabiei</i>	G1535D	IIIS6		Yes	
		<i>Rhipicephalus microplus</i>	F1538I	IIIS6		Yes	
		<i>Panonychus citri</i>	F1538I	IIIS6		Yes	
		<i>Helicoverpa zea</i> <i>Heliothis virescens</i>	D1549, E1553G	IIIS6-IVS1		Yes	
4	Nicotinic acetylcholine receptor	<i>Myzus persicae</i>	R81T	$\beta$		Yes	<a href="#">Bass et al. (2011) BMC Neuroscience. 12:51</a> <a href="#">Beckingham et al. (2013) Pest Biochem Phys. 107:293</a> <a href="#">Panini et al. (2014) Pest Manag Sci. 70:931</a> <a href="#">Puinean et al. (2013) Pest Manag Sci. 69:195</a> <a href="#">Koo et al. (2014) Crop Protection. 55:91</a> <a href="#">Kim et al. (2015) J Asia-Pacific Entomol. 18:291</a> <a href="#">Liu et al. (2005) PNAS. 102:8420</a>
5	Nicotinic acetylcholine receptor	<i>Aphis gossypii</i>	R81T	$\beta$		Yes	
		<i>Nilaparvata lugens</i>	Y151S			No	
6	Chloride channel	<i>Tetranychus urticae</i>	G323D			Yes	<a href="#">Kwon et al. (2010) Insect Mol Biol. 19:583</a> <a href="#">Ilis et al. (2014) Insect Biochem Mol Biol. 48:17</a> <a href="#">Wang et al. (2015) Insect Mol Biol. 25:116</a>
		<i>Plutella xylostella</i>	G314D/G323D, G326E			Yes	
		<i>Frankliniella occidentalis</i>					
		<i>Thrips palmi</i>	G275E	$\alpha 6$		Yes	
		<i>Drosophila melanogaster</i>	P146S	$\alpha 6$		No	
10	Chitin synthase (CHS1)	<i>Tetranychus urticae</i>	I1017F			Yes	<a href="#">Van Leeuwen et al. (2012) PNAS. 109:4407</a> <a href="#">Ilis et al. (2014) Insect Biochem Mol Biol. 48:17</a> <a href="#">Demaeest et al. (2014) Insect Biochem Mol Biol. 51:52</a> <a href="#">Dourie et al. (2016) PNAS. 113:14692</a>
11	Microbial disruptors of insect midgut membranes	multiple	multiple			Yes	<a href="#">Xiao, Y., and K. Wu (2019) Philos Trans R Soc Lond B Biol Sci. 374(1767): 20180316.</a> <a href="#">Tabashnik, B. E., and Y. Carrere (2020) J Econ Entomol. 113(2): 553-561.</a>
15	Chitin synthase (CHS1)	<i>Culex pipiens</i>	I1043L/M I1043F			Yes	<a href="#">Poretta et al (2019) Acta Tropica. 193:106</a> <a href="#">Fotakis et al (2020) PLOS Negl Trop Dis. 14:e0008284</a>
20	Mitochondrial Complex III electron transport	<i>Tetranychus urticae</i>	G126S, G132A, A133T, I136T, S141F, P262T			Yes	<a href="#">Van Leeuwen et al. (2008) PNAS. 105:5980</a> <a href="#">Van Nieuwenhuysse et al. (2008) Pest Manag Sci. 65:404</a> <a href="#">Ilis et al. (2014) Insect Biochem Mol Biol. 48:17</a> <a href="#">Fotoukiani et al. (2020) Pest Manag Sci.</a>
22	Voltage-dependent sodium channel blockers	<i>Plutella xylostella</i>	F1845Y, V1848I			Yes	<a href="#">Wang et al. (2015) Insect Sci. 23:50</a> <a href="#">Samantidis et al. (2019) Insect Biochem Mol Biol. 104:73</a>
23	Acetyl-CoA carboxylase	<i>Trialeurodes vaporariorum</i> <i>Bemisia tabaci</i>	E645K A2083V			No Yes	<a href="#">Karatolos et al. (2012) Insect Mol Biol. 21:327</a> <a href="#">Lueke et al. (2020) Pest Biochem Mol Biol</a>
28	Ryanodine receptor	<i>Plutella xylostella</i> <i>Tuta absoluta</i> <i>Chilo suppressalis</i> <i>Spodoptera frugiperda</i> <i>Spodoptera exigua</i> <i>Chilo suppressalis</i> <i>Plutella xylostella, Tuta absoluta, C. suppressalis, Spodoptera frugiperda, Spodoptera exigua</i>	G4946E E1338D, Q4549L, I4790M G4903E/V, I4746M G4910E I4734M I4743M I4758M, Y4667D/C, G4915E, Y4891F			Yes Yes Yes Yes Yes Yes Yes	<a href="#">Troczka et al. (2012) Insect Biochem Mol Biol. 42:873</a> <a href="#">Guo et al. (2014) Scientific Reports. 4:6924</a> <a href="#">Roditiakis et al. (2017) Insect Biochem Mol Biol. 80:11</a> <a href="http://onlinelibrary.wiley.com/doi/10.1002/ps.4439/pdf">http://onlinelibrary.wiley.com/doi/10.1002/ps.4439/pdf</a> <a href="#">Boaventura, D., et al. (2020) Pest Management Science. 76(1): 47-54.</a> <a href="#">Zuo, Y. Y., et al. (2019) Insect Sci.</a> <a href="#">Huang, J.-M., et al. (2020) Insect Biochem Mol Biol. 121:103367</a> <a href="#">Richardson, E. B., et al. (2020) Journal of Pest Science.</a>

Mutations with "No" listed under "Field Relevance" indicates resistance was observed under laboratory conditions though not currently identified in the field.